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Multi-axial non-contact in situ micromanipulation by steady streaming around two oscillating cylinders on holonomic miniature robots

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Abstract

In the field of micromanipulation, an in situ three-axial rotation of a microscale object remains difficult to realize, with rotational resolution and repeatability remaining low. In this paper, we describe the fundamental principle, properties, and experimental results of multi-axial non-contact in situ micromanipulation of an egg cell driven by steady streaming generated around an oscillating cylinder. A continuously oscillating cylinder generates the steady streaming that draws an egg cell toward the cylinder. If it is trapped by an eddy near the tip of the cylinder, it continuously rotates around the vertical axis at a fixed point. If it is trapped by a swirl flow generated around the side of the cylinder, it rotates around the horizontal axis. We define Reynolds number, R_e , as $ar_c\omega/\nu$, where a is half of the oscillation's amplitude, r_c is the cylinder's radius, ω is the oscillation's angular frequency, and ν is the kinematic viscosity. We demonstrate that the conditions of the vertical and horizontal rotations are determined by two dimensionless numbers: R_e and a/r_c . In our experiments, we obtained rotational resolutions of 0.05° and 0.11° and maximal angular velocities of 34.8°/s and 188°/s for the vertical and horizontal rotations, respectively. We also developed unique micromanipulation methods using two oscillating pipettes attached to holonomic miniature robots. We successfully manipulated five degrees of freedom (DoF) of the cell (three posture angles and two translational displacements along the X and Y axes) with the steady streaming. The proposed method enables a multi-axial, non-contact, in situ, and compact micromanipulation independent of the electrical, optical, magnetic, shape, and stiffness properties of the objects; moreover, it can be applied in microfluidics, biomedical, and heterogeneous microassembly applications.

Keywords In situ multi-axial micromanipulation · Steady streaming · Oscillating cylinder · Holonomic miniature robot

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1 Introduction

Recently, micromanipulation ranging from 1 μ m to 1 mm has generated significant interest (Savia and Koivo 2009; Cecil et al. 2016). Micromanipulations can be categorized as air, vacuum, and liquid, according to the medium in which the microscopic objects are manipulated (Aoyama and Fuchiwaki 2001; Fuchiwaki et al. 2008; Xie and Regnier 2011; Rong et al. 2014).

The micromanipulation in air is important for surface mounting technology (SMT) of electronic chip parts. Owing to a rapid growth in the demand for smaller personal computers, there is an increasing need to reduce the size of electronic components. However, three main problems are associated with micromanipulation in air. One is the "sticky problem;" microscopic objects can be straightforwardly trapped owing to the action of adhesive forces such as the electrostatic force, interfacial force, and/or viscous drag force, all of which are generally negligible in manipulations of supra-cm-size objects. The second problem is the challenge of three-axial rotation; it is difficult to control three posture angles of a small object by fixing its central point because manipulators become larger in size and heavy if three rotational stages are added (Hatamura et al. 1995). With mobile precise robots for positioning the micromanipulators, manipulation becomes relatively compact and flexible (Fatikow et al. 2000; Fuchiwaki et al. 2012, 2014), although these mobile robots should be rotated precisely around the central point of the manipulated microscopic object (Fuchiwaki et al. 2015). The third problem is associated with the limitation of the shape of the object; the shape should be cubic because it is convenient to fix two posture angles of a cube around two horizontal axes by putting them on their base plate (Chen et al. 2010; Zhang et al. 2010; Tamadazte et al. 2011; Boudaoud 2012). However, the smaller the objects become, the higher the risk of damaging them or flicking them away by conventional manipulations. These problems can be alleviated using the liquid's capillary force for micromanipulation (Obata et al. 2004; Kumagai and Fuchiwaki 2012; Fuchiwaki and Kumagai 2013; Fan et al. 2015). However, regulating the capillary force remains challenging because small droplets of water (below 1 mm in size) dry rapidly, and most non-drying liquids introduce contaminations. These problems intensify when micromanipulating complex-shaped and fragile objects.

Micromanipulation in a liquid significantly alleviates these three problems because a liquid flow carries and rotates the immersed microscopic objects and protects them by the liquid layer. Most of the sub-1-mm-size living creatures dwell in water because it is their optimal dwelling medium. Blood vessels are highly effective systems for transportation of nutrients (Hyakutake et al. 2015; Hyakutake and Nagai 2015). When we use grippers for grabbing fragile objects such as biological cells in a liquid, we need to precisely regulate the grasping force; otherwise, the grasped cells will be damaged (Kim et al. 2008; Yabugaki et al. 2013). A significant amount of research has been performed on developing methods for indirect micromanipulation of biological cells in liquids, using optical tweezers (Grier 2003; Chen et al. 2013; Ozkan et al. 2003; Shafiee et al. 2009; Chowdhury et al. 2013), although optical property of the targets affects their manipulating properties. Particle-based micromanipulation by optical or magnetic forces has also been categorized as contact manipulation in liquids (Gijs 2004; Yesin et al. 2006; Floyd et al. 2009; Elbuken et al. 2009). Those techniques can still damage fragile objects.

Acoustic streaming is an interesting phenomenon because it exhibits steady-streaming patterns that change according to the frequency and amplitude of oscillations (Tatsuno 1973, 1981; Wang and Drachman 1982; Sadhai 2012; Nuriev and Zaitseva 2014; Coenen 2016). Recently,

we have proposed and studied a method and applications for steady-streaming-based micromanipulation that utilizes an oscillating cylinder (Misaki et al. 2004, 2007; US Patent 2005; Aoyama 2006). The steady streaming is among the preferable methods of non-contact micromanipulation because objects can be manipulated independent of their electrical, optical, magnetic, shape, surface, and stiffness properties. Significant attention has been devoted to this issue (Lieu 2012; Hattori et al. 2015; Leibacher 2015; Amit et al. 2016). Combining acoustic streaming and microscale channels is another potential option for selective transportation of microscale objects (Jeong et al. 2011; Büyükkoçak et al. 2014; Hayakawa et al. 2014; Hayakawa 2015). Several interesting studies have addressed the issue of micromanipulation using acoustic streaming (Friend and Yeo 2011; Walter et al. 2012; Wiklund et al. 2012; Destgeera and Sung 2015).

In the biomedical field, micromanipulation of egg cells is important for realizing transgenic animals and microscale fertilization. No contact micromanipulation using AC-field (Benhal 2014), hydrodynamic flow (Yalikun et al. 2016a, b; Leung et al. 2012), magnetic field-driven rotating particle (Ye et al. 2012) methods are also feasible methods for fragile biomedical cells. In supporting information, Table 1 shows the comparison of those methods with the proposed method for the particle of ~100 µm in diameter. AC-field method (Benhal et al. 2014) has 0.57° of rotational resolution in vertical rotation, although electrical property of the target affects their manipulating property. The hydrodynamic flow method (Yalikun et al. 2016a, b) has advantage for the fast and powerful micromanipulation. The magnetic field-driven rotating particle method (Ye et al. 2012) are usable in closed narrow chamber. However, the in situ three-axial rotation still remains to achieve using these methods, with rotational resolution and repeatability remaining low.

The motivations of this study are to experimentally realize the precise multi-axial in situ rotation of a micro-object by fixing its central point, and to investigate the specifications thorough evaluation of the experimental results from fluid dynamic aspects. The development of a compact manipulating method to demonstrate its feasibility is also a motivation. In this paper, we describe the fundamental principle, properties, and experimental results of the multi-axial micromanipulation of egg cells driven by the steady streaming generated around an oscillating pipette attached immediately above the bottom of a Petri dish. When the cylinder oscillates continuously, the generated acoustic stream draws the microscopic object toward the cylinder. If it is trapped by an eddy in front of the tip of the cylinder, it rotates around the vertical axis. If it is trapped by a swirl flow generated around the side of the cylinder, it rotates around the horizontal axis. For up to one oscillation cycle of the cylinder, the slight eddy or the slight swirl rotates it by a marginal rotational angle. In the experiments, we realized non-contact drawing, trapping, and two-axial rotation around the vertical and horizontal axes of the egg cell. In Sect. 2, we explain the condition and the experimental setup. In Sect. 3, we classify the streaming by fluid structures. In Sect. 4, we describe the fundamental performances of the vertical and horizontal rotations and the drawing. In Sect. 5, we describe multi-axial in situ microinjection assisted by oscillating pipettes positioned by holonomic miniature robots (Fuchiwaki 2013) to demonstrate the feasibility. In Sect. 6, we discuss the potential applications in microfluidics and microassembly.

2 Parameters and experimental setup

2.1 Parameters and conditions

The Reynolds number, R_{e} , is defined as

$$R_{\rm e} = \frac{r_{\rm c} \cdot a\omega}{v},\tag{1}$$

where *a* is half of the oscillation's amplitude, r_c is the cylinder's radius, ω is the oscillation's angular frequency, and ν is the kinematic viscosity. The cylinder's radius r_c is the characteristic length, and the quantity $a\omega$ is the characteristic velocity. The ratio a/r_c is an important dimensionless parameter that determines the flow patterns as well as R_e (Tatsuno 1973). An example of an acoustic stream around an oscillating straight cylinder is shown in Fig. 1 (Tatsuno 1981). In the caption of Fig. 1, *f* is the oscillation's frequency. The double arrow indicates the direction of the cylinder's oscillation. The dotted line indicates the bottom of the Petri dish in the present work. Symmetrical circulatory streaming toward



Fig. 1 Image of trajectories of microscopic glass particles with diameters of ~10 µm, with $\nu = 4.15 \times 10^{-5} \text{ m}^2/\text{s}$, $R_e = 4.61$, $a/r_c = 0.33$, f = 10 Hz, $r_c = 3020 \text{ µm}$, a = 1009 µm, and exposure time of 10 s. The dotted line indicates the bottom of the container in the present work. Reproduced from ref. Tatsuno (1981) with permission

the cylinder along the axis of the oscillation and away from the cylinder perpendicular to the axis is evident. In the present work, we replaced the straight cylinder with a glass pipette that was bent at 40° and attached immediately above the bottom of the Petri dish filled with pure water. In Sect. 3, we performed experiments to categorize the flow patterns of the acoustic streaming generated by the oscillating bent cylinder, in terms of two dimensionless parameters, R_e and a/r_c . In this study, we focused on the micromanipulation of frozen-thawed fertilized egg cells of mice for $r_e/r_c = \sim 1.2$, with $r_c = 40 \ \mu$ m. Here, r_e is defined as the radius of the manipulated egg cell.

2.2 Experimental setup

Figure 2a schematically shows the oscillating cylinder driven by a piezoelectric actuator. In the supporting information, Table 2 lists the performance characteristics of the actuator. We denote the oscillating axis as Y_p , the horizontal axis perpendicular to Y_p as X_p , and the vertical axis as Z_p . We used a pipette with an outer diameter of 80 µm as the oscillating cylinder. The pipette was bent at 40° to align it parallel to the bottom. Figure 2b shows the experimental setup for observing the steady streaming in the X_pY_p plane. The oscillating pipette was attached to an *XYZ* stage (Eppendorf, 5171). The *XYZ* stage was attached to an inverted microscope (Olympus, IX-71). A microscopic image was captured by a high-speed camera (PHOTRON, FAST-CAM NET-C), with the rate of



(b) Observing the flow in $X_P Y_P$ plane using inverted microscopy.

Fig. 2 Experimental setup

image-capture reaching 250 fps, through the bottom of the Petri dish.

The pipette was attached immediately above the bottom, without contact. The oscillation of the pipette was a sinusoidal wave with the half amplitude displacement ranging from 2 to 25 μ m. The displacement was approximately proportional to the applied voltage, although it was affected by the hysteresis-related nonlinearity of the piezoelectric actuator. It was also affected by the viscosity of the liquid and the mechanical resonance frequency; thus, we adjusted the displacement by varying the voltage.

We measured the displacement by the pipette's residual images captured using the fast-speed camera. The microscope's resolution was 0.6 μ m; the measurement errors for the X and Y axes was estimated \pm 0.3 μ m. The focal depth was 2.9 μ m; the error for the Z-axis was estimated \pm 1.5 μ m. We varied the oscillation frequency from 10 to 250 Hz.

In the experimental work described Sect. 3, a few drops of Indian ink were added as tracer particles into the Petri dish filled with pure water, to investigate the flow patterns. In the work described in Sects. 4 and 5, we put the frozen–thawed fertilized egg cells of mice into the Petri dish filled with a culture solution of M2 to investigate the fundamental manipulating performances. In Sect. 5.2, we provide additional explanation about the use of the holonomic miniature robots for positioning additional oscillating cylinder.

3 Flow patterns

3.1 Condition of steady streaming

Figure 3a shows the diagram of the flow state around the cylinder in the R_e - a/r_c plane for $R_e < 1$ and $a/r_c < 0.64$. When $R_e > 0.12$, we observed the steady streaming. When $R_e < 0.08$, the tracer particles oscillated around their initial positions. Although more detailed experimental analysis is necessary to demonstrate the occurrence of the transition with R_e , we estimated the transition area to be from 0.08 to 0.12 of R_e for the tracer particles to flow along the streaming.

3.2 Classification by fluid structure

Figure 3b shows the typical trajectories of the tracer particles, plotted every 0.1 s during the window of 0.8 s, with $(R_e, a/r_c) = (0.49, 0.40)$. For R_e in the range 0.12–1.0, symmetrical steady vertical twin eddies were generated in front



layer of the inner circulatory streaming.

Fig. 3 Experimental results of steady-streaming patterns

of the pipette in the $X_P Y_P$ plane. We hereby call them "vertical eddies".

Symmetrical backward and forward curved flows were also generated around the sides of the pipette in Fig. 3b. We assume that the observed curved flows correspond to the parts of horizontal swirl flows because the flows are approximately half of the axisymmetric circulatory streaming in Fig. 1, if we consider the dotted line to be the bottom of the container. We hereby call them "horizontal swirl flows."

We also observed two types of steady streaming toward the edges and the sides of the cylinder in Fig. 3b. Those two flows correspond to the parts of the streaming toward the cylinder along the axis of the oscillation in Fig. 1.

Figure 3c shows the classification of the steady streaming by fluid structures and the definitions of the symbols and colors in Fig. 3b, d. We hereby classify "vertical eddies" and "horizontal swirl flows" as "inner circulatory streaming" and the other two streaming as "outer streaming". The inner circulatory streaming and the outer streaming are expected to rotate and draw microscopic objects, respectively.

3.3 Layer of inner circulatory streaming

To investigate the shape of the layer of the inner circulatory streaming, we introduced smaller tracer particles of Indian ink into the liquid. Figure 3d shows two typical images of their distribution with $(R_e, a/r_c) = (0.57, 0.30)$ and (0.42, 0.90). The layer was keyhole-shaped, and its thickness varied from 20 to 200 µm from the outline of the cylinder.

4 Fundamental manipulating performances

4.1 Vertical rotation

Figure 4a schematically shows the flow pattern of the vertical rotation. Figure 4b shows the motion of the studied egg cell in plots of a/r_c vs. R_e . To exhibit the flow patterns without it, the results in Fig. 3a were replotted in Fig. 4b. Three motion patterns are observed in Fig. 4b. For R_e below 0.07, it oscillated around its initial position (Mode i). For R_e above 0.07, it rotated around the vertical axis while fixing the position of the axis (Mode ii). However, for a/r_c above 0.2 and R_e above 0.25, it was lifted from the bottom and then rolled in the $-X_p$ direction (Mode iii). In the supporting information, Table 3 lists the properties of the vertical rotation without the rolling. Figure 4c shows snapshots of the two rotation modes.

Figure 4d shows the two typical trajectories of the central point with a vertical rotation, with $(R_e, a/r_c) = (0.09, 0.15)$ and (0.27, 0.11). The pair of coordinates (X_c, Y_c) is the position of the average rotational axis. Figure 4e shows plots of each of ω_z , $\Delta \theta_z$, and $L_{xy} \pm \delta r_y$ vs. R_e . Here, ω_z is the angular

velocity. $\Delta \theta_{z}$ is the rotational resolution during a cycle of the cylinder's oscillation. L_{xy} is the distance between O and (X_c, Y_c) . δr_v is the radial run out and is determined by the maximal distance between (X_c, Y_c) and the central point. From the left panel of Fig. 4e, ω_{z} is approximately proportional to $R_{\rm e}$. The maximal value of ω_z is 34.8°/s for ($R_{\rm e}$, $a/r_{\rm c}$) = (0.27, 0.11). From the middle panel of Fig. 4e, $\Delta \theta_{\rm z}$ is approximately proportional to R_e , and a smaller a/r_c is effective for decreasing $\Delta \theta_z$. The minimal value of $\Delta \theta_z$ is 0.05° for $(R_e, a/r_c) = (0.12, 0.05)$. Because of the low R_e , it stops immediately after the oscillation stops. This is advantageous for the capability to precisely regulate the rotational angles. In the right panel of Fig. 4e, the error bars are the corresponding δr_{v} . We observe that both L_{XY} and δr_{v} decrease as $R_{\rm e}$ increases. We also observe similar characteristics for the form of the two trajectories in Fig. 4d.

For precise and rapid microscopic operations, smaller a/r_c and higher R_e are feasible, owing to the smaller $\Delta \theta_z$ and δr_v and higher ω_z . At these values of $(R_e, a/r_c)$, (0.27, 0.11) and (0.12, 0.05) should be selected for coarse and precise vertical rotations, respectively.

4.2 Horizontal rotation

Figure 5a schematically shows the flow pattern for the horizontal rotation. Figure 5b shows the motion patterns for both the vertical and horizontal rotations in the plots of a/r_c vs. R_e .

Four motion patterns are observed for the horizontal rotation in Fig. 5b. For $R_{\rm e}$ below ~0.07, the egg cell oscillated around its initial position (Mode I). For $R_{\rm e}$ in the range 0.07-0.25, it oscillated and was in contact with the cylinder because the horizontal swirl flow was not sufficiently rapid to lift it from the bottom (Mode II). For R_e above 0.25, it rotated around the horizontal axis because the swirl flow became sufficiently rapid to lift it, although the horizontal translation toward the $-X_p$ direction occurred simultaneously (Modes III and IV). The horizontal rotation requires larger $R_{\rm e}$ than the vertical rotation does because additional energy is required for lifting it from the bottom. If excessive energy is supplied to the vertical rotation, e.g., if $a/r_{\rm c}$ is larger than ~ 0.2 and R_e is larger than ~ 0.27, it levitates, starts to roll, and then screws in the $-X_{\rm P}$ direction (Mode III-IV). Figure 5c shows snapshots of the contact oscillation and the horizontal rotation.

Figure 5d shows the typical range of the horizontal rotation. $X_{c_{-l}}$ and $X_{c_{-u}}$ are the lower and upper boundaries, respectively, of X_c . ΔX_c is the length between the boundaries, defined by $|X_{c_{-u}} - X_{c_{-l}}|$. The egg cell does not rotate outside this range. We posited that the tapering and the bent shape of the oscillating pipette provide the lower limit of $X_{c_{-l}}$, although the upper limit of $X_{c_{-u}}$ is determined by the interaction of the vertical eddy and the horizontal swirl flow.



Fig. 4 Experimental results of vertical rotations

Figure 5e shows the plots of each of ω_h , $|V_x|$, and $L_{YZ} \pm \delta r_h$ vs. R_e . Here, ω_h is the angular velocity. V_x is the velocity in the X direction. L_{YZ} is the distance between $(X_c, 0, 0)$ and (X_c, Y_c, Z_c) . δr_h is defined as the radial run out of the horizontal rotation. From the left panel of Fig. 5e, ω_h is approximately proportional to R_e as well as ω_z . The maximum of ω_h is 188°/s at $(R_e, a/r_c) = (1.03, 0.43)$. From the middle panel of Fig. 5e, $|V_x|$ is approximately proportional

to $R_{\rm e}$. From the right panel of Fig. 5e, L_{YZ} decreases for higher $R_{\rm e}$ as well as L_{XY} , and $\delta r_{\rm h}$ was below 10 µm. This verifies that it does not contact the cylinder. In the supporting information, Table 4 lists the properties of the noncontact horizontal rotation. As shown in Table 4, $\Delta \theta_{\rm h}$ is the rotational resolution during a cycle of the cylinder's oscillation. The minimum of $\Delta \theta_{\rm h}$ is 0.11° for ($R_{\rm e}$, $a/r_{\rm c}$) = (0.28, 0.18).



Fig. 5 Experimental results of horizontal rotations

Although smaller $\Delta \theta_{\rm h}$ and higher $\omega_{\rm h}$ are important for precise and rapid microscopic operations, decreasing $|V_x|$ is also important because the horizontal translation

is undesirable for regulating the rotational angle. We concluded that the region with $R_e = 0.25-0.5$ and $a/r_c =$

0.1-0.2 (Mode III in Fig. 5b) is appropriate for the horizontal rotation.

4.3 Drawing

Figure 6a schematically shows the flow pattern for the drawing. Figure 6b shows the typical trajectories from various initial positions along the $Y_{\rm P}$ axis for $(R_e, a/r_c) = (0.68,$ 0.42). Here, we denote the initial position as (X_i, Y_i) . It shifts to the tip of the cylinder after it reaches the side when Y_i is in the range 200–350 μ m, although it shifts in the – $X_{\rm p}$ direction when Y_i is in the range 410–480 µm; this is because the range of the horizontal rotation is $X_{\rm P} < -134 \,\mu{\rm m}$, as shown in Fig. 5d. Figure 6c shows the plots of t_d vs. Y_i . Here, t_d is the drawing time from Y_i to Y_c . If we estimate that Y_i logarithmically depends on t_d , t_d is 25 s when $Y_i =$ 500 μ m. $R_{\rm e}$ should be increased for widening the range and shortening t_d .

5 Multi-axial in situ micromanipulation

5.1 Conventional method

Two methods can be employed to rotate the egg cell using a holding and an injection pipettes, as shown in Fig. 7a. One amounts to randomly rotating it by ejecting and suctioning the liquid inside the holding pipette, as shown in the top panel. Another method amounts to scratching it using the injection pipette with weak suctioning by the holding pipette, as shown in the bottom panel. It is generally challenging to implement either of these methods, particularly by inexperienced personnel. In this chapter, we propose a more straightforward, safer, and more precise micromanipulation based on the steady streaming.

5.2 An oscillating cylinder

To determine the feasibility of the proposed method, we conducted microinjection supported by the oscillation of the holding pipette. Figure 7b shows the trajectories of the drawing and two-axial rotations by the steady streaming. Figure 7c shows snapshots of the microinjection process. First, we positioned it near the holding pipette by drawing. Then, we rotated it around the horizontal axis until its polar body was in the focus plane. Next, we rotated it around the vertical axis until its polar body pointed downward or upward to prevent damage to the polar body by the injection pipette. Then, we captured it by suctioning the liquid using the holding pipette and injected the culture solution of M2 into the nuclei of five frozen-thawed fertilized eggs of mice to investigate the effect of the proposed method on the cells' differentiation rate.

In the supporting information, figure S1 shows the photograph of the fertilized egg cells after injection of the culture solution assisted by the steady streaming around the oscillating holding pipette. We verified that four out of the five egg cells underwent differentiation. Because the differentiation rate of frozen-thawed oocytes was reported to be 75-95% depending on the oocytes and experimental conditions



(b) Relationship between initial position and trajectory.



Fig. 7 Microinjection assisted by the steady streaming around an oscillating holding pipette

(freezing and thawing methods, culture solution, and the operator's techniques) (Nakagata 1989; Carroll et al. 1990; Borini et al. 2006; Bianchi et al. 2007), we did not find the influence of the proposed method on the differentiation rate.

The flows draw, trap, and rotate the egg cell around the vertical and horizontal axis sequentially; thus, we change R_e and a/r_c (f and a) according to the position and desired manipulation by referring to the diagram in Fig. 5b and trajectories in Fig. 6b. However, during this operation, we were required to shift the holding pipette between the horizontal and rotational axes; the efficiency can be potentially increased, although additional oscillating pipettes are required.

5.3 Two oscillating cylinders

In this section, we describe a method for more efficient multi-axial in situ micromanipulation using two additional oscillating pipettes. Figure 8a shows an image of the setup for a microinjection. We added two holonomic miniature robots with manual Z-axis stages for positioning the oscillating pipettes to make the manipulation compact (Fuchiwaki 2013).

Figure 8b shows the trajectories of drawing toward the intersection point of the two rotational axes by combining the two oscillating pipettes. This special arrangement allows us to draw and rotate all three of the posture angles sequentially, fixing the egg cell's central point without any contact. We implemented five degrees of freedom (DoF) noncontact manipulation of the egg cell (three posture angles and two translational displacements in X and Y axes) with the angular resolution better than 0.05° and 0.11°, and the radial run out of 4 and 10 µm for the vertical and horizontal rotations, respectively. Because the oscillating pipettes were attached to the holonomic robots that can move X, Y, and Θ -axes independently with positioning resolution better than 10 nm and 10^{-5} degrees, we were able to change the position and the posture of the pipettes; we were able to change the intersection point of the two rotational axes and the drawing directions, flexibly, and precisely.

Figure 8c shows snapshots of the microinjection process. There are two working layers. The bottom layer is for the drawing and for the horizontal and the vertical rotations by two additional oscillating pipettes. The top layer is for the microinjection using the conventional holding and injection pipettes. First, we positioned the cell near the two oscillating pipettes and the holding pipette in the bottom layer by



rotations of the egg cell by two oscillating cylinders

two additional oscillating pipettes.

Fig.8 Multi-axial non-contact in situ micromanipulation by the steady streaming around a pair of oscillating cylinders

drawing. Next, we rotated it around the horizontal axis until its polar body was in the focus plane. Then, we rotated it around the vertical axis until its polar body pointed downward or upward. Finally, we captured it using the holding pipette and lifted the cell to the top layer for performing the injection.

The average processing time required by four inexperienced personnel was 15 s, although the same task required 60–300 s when using the conventional methods in Fig. 7a. Thus, the proposed method can improve the processing time, particularly for inexperienced personnel.

5.4 Discussion

In the supporting information, Table 5 shows the specification of the multi-axial non-contact in situ micromanipulation by the steady streaming around the two oscillating cylinders on the holonomic robots for $r_e/r_c = \sim 1.2$. We also verified that glass and alginate spheres of diameters ~ 100 µm could be rotated around the vertical and horizontal axes and could be drawn to the pipette; the characteristics of their manipulation were similar to those of the studied egg cells. Therefore, we consider the elasticity of the targets does not give a large influence on the rotational resolution and the angular velocity, although more detailed evaluation is required. Although investigation of the operating range of the size of particles is also one of the future subjects, we estimate that sub-millimeter to the sub-micro-meter-sized particles are able to manipulated with similar characteristics under the similar conditions of R_e , a/r_c , and r_e/r_c from the similarity law of hydrodynamics. The lower limitation should be determined by the Brownian motion. To determine the upper limitation, we should consider the influence of the gravity force, buoyancy, and inertia force on the manipulating objects

because those volume-proportional forces are not negligible compared with the surface-proportional viscous drag force generated by the steady streaming for supra-mm-sized objects. The oscillating cylinder-based fluidic manipulation is feasible for multi-axial non-contact in situ manipulation of micro-objects with different shapes, rigidity, magnetic and optical properties.

6 Summary and future prospects

We described the basic principle, characteristics, and experimental results of the multi-axial micromanipulation method based on the steady streaming generated around an oscillating cylinder. In our experiments, we precisely rotated an egg cell around vertical and horizontal axes. We showed that the conditions of the rotations are determined by R_{e} and the ratio of a half of the amplitude to the radius of the cylinder, a/r_c . The minimal angular resolutions were 0.05° and 0.11° and the maximal angular velocities were 34.8°/s and 188°/s for the vertical and horizontal rotations, respectively. We also investigated the range and required time for drawing it to the cylinder. To demonstrate the feasibility, we conducted microinjection using the oscillating pipette. We were able to control its five DoF, consisting of three posture angles and two translational displacements in X and Y axes, without any contact.

We also explain the special arrangement of the two oscillating pipettes on holonomic miniature robots. The arrangement permitted us to draw it toward the point of intersection of the two rotational axes and also to regulate its three posture angles sequentially by fixing the central point. The average processing time of microinjection by four inexperienced personnel decreased to 15 s from 60 to 300 s.

The proposed method enables a delicate, multi-axial, noncontact, in situ, and compact micromanipulation independent of the electrical, optical, magnetic, shape, and stiffness properties of the microscopic objects and which can be used in microfluidics, biomedical, and microassembly applications. Although investigation of the operating range of the size of particles is one of the future subjects, we estimate that sub-milli-meter to sub-micro-meter-sized heterogeneous particles could also be manipulated with similar characteristics under the similar conditions of R_e , a/r_c , and r_e/r_c from the similarity law of hydrodynamics.

Future work will aim at tailoring this microscopic manipulation method to applications in microfluidics, biomedicine, and microscale assembly.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

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